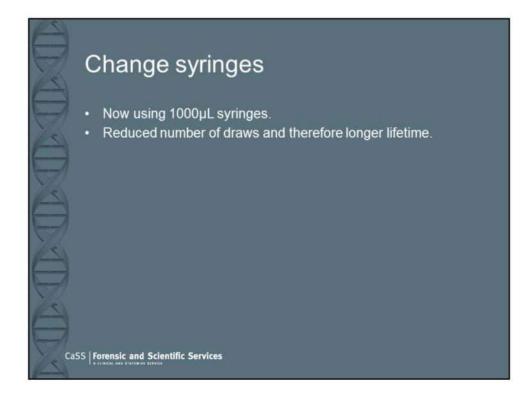


Outline • Automated DNA IQ enhancements • Testing results so far • Outcome summary from external audit

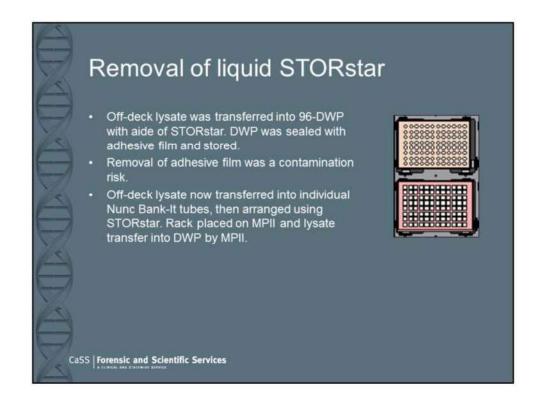
Automated DNA IQ enhancements

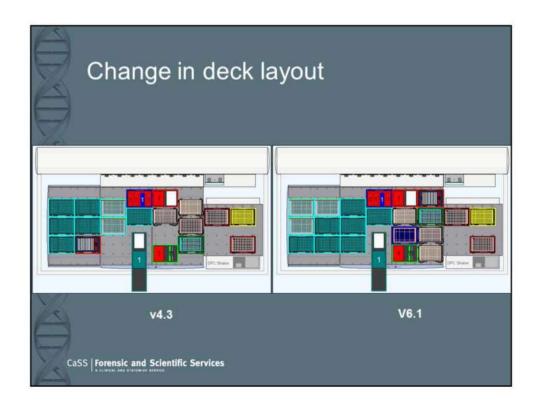
- 1. Change syringes from 500µL to 1000µL
- 2. Change in off-deck lysis volume from 500µL to 300µL
- 3. Removal of liquid STORstar
- 4. Change of deck layout
- 5. Automated addition of DNA IQ Resin
- 6. Off-board mixing of DNA IQ Resin
- 7. Magnet changed from PKI to ABI
- 8. Electronic platemap changed for volumes & new steps
- 9. Risks considered with regards to droplets on side of tips
- 10. Changes reviewed

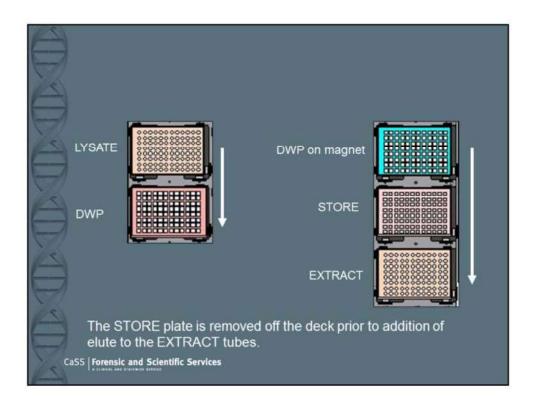


Change in off-deck lysis volume

- Volume of extract in well/tube is reduced to minimise contamination risk during shaking/incubating.
- Translates also to a reduction in reagent usage.
- · Minimal difference in yield (as found from QPS Tapelift Trial).





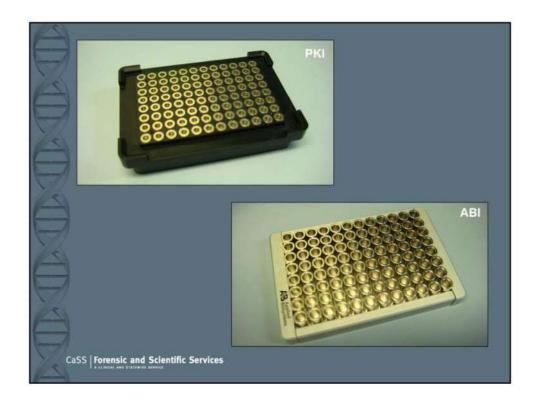


Automated addition/mixing of resin

- Resin was added manually and then mixed by MPII.
- Resin now added by MPII, followed by addition of lysis buffer (no mixing).
- User intervention required: seal plate with septa mat and shake on MixMate at 1100rpm for 5mins followed by centrifugation at 3000rpm for 2mins to bring down any drops.
- · Septa mat is removed and DWP returned to MPII.
- This mixing is KEY to improving the DNA recovery.

Magnet changed

- PKI magnet had corners that required the user to "click in" the plate. If the plate is not straight, resin loss can occur.
- ABI magnet has no corners and the magnets are raised, so the plates sit better and no resin loss occurs.



Platemap changed

- Electronic platemap for MPII has changed to reflect new pipetting volumes.
- The addition of the Nunc "lysate" rack from off-deck lysis meant the addition of 1 new column of information in the platemap.

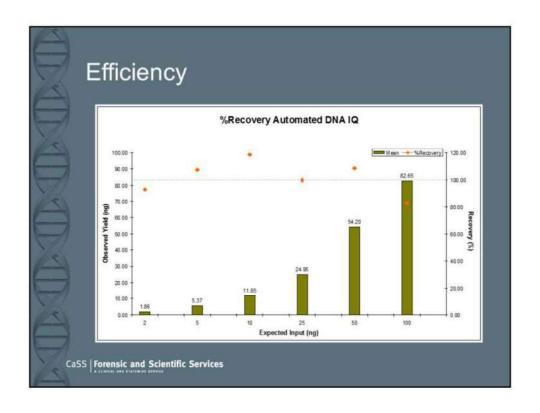
Risks considered

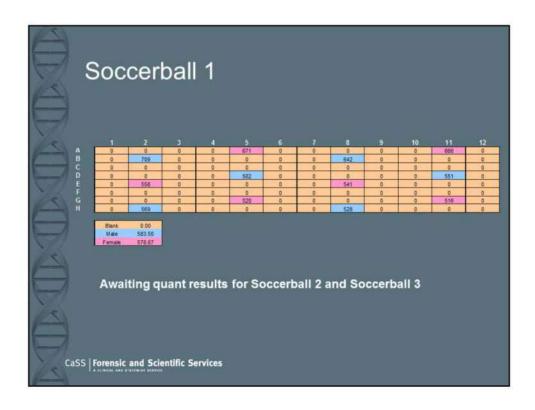
- Static attraction of the tip causes some reagent types (e.g. lysis buffer) to adhere to the tip side.
- These are only reagents and the risk of sample contamination is very low.
- Pre-step, post-step and transport air gaps were optimised to make the pipetting cleaner.

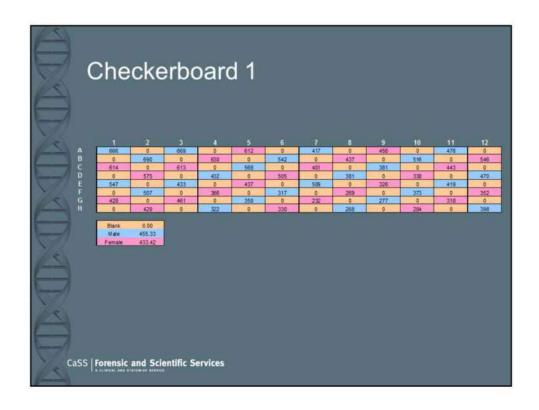


Testing results so far

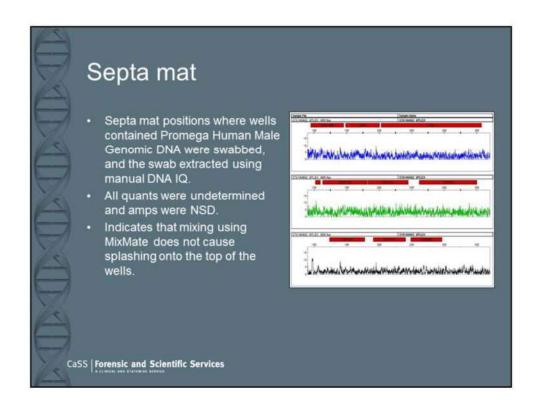
- 1 efficiency plate
- · 3 soccerball plates
- 1 checkerboard plate
- · 1 zebra stripe plate
- 96-sample plate in progress
- The combination of these plates provide crucial information











External audit

- · Assoc. Prof Theo Sloots and David Whiley
- · They will write a summary report of their observations.
 - The adhesive seal that we had used on the off-deck lysis plate was a problem. This process is no longer in the current version of DNA IQ.
 - The MP II and current version of the automated DNA IQ is OK.
 - Commended us on our environmental monitoring and monthly cleaning regimes.
 - Commended us on our thorough approach to identifying and diagnosing the problems.
 - We're "ahead of the pack" and learning lessons that nobody else has come across yet.

Conclusion

- Enhancements have been made to the MP II. These changes have been tested and approved by a PerkinElmer LHS.
- The changes have caused an increase in efficiency and recovery of DNA.
- Anti-contamination checks have been run and results are as expected.
- · Feedback from external auditors were very positive.